

# VirusGEN® LV Transfection Kit

## Quick Reference Protocol

Instructions for MIR 6760

Full protocol, SDS and Certificate of Analysis available at [mirusbio.com/virusgen](http://mirusbio.com/virusgen)



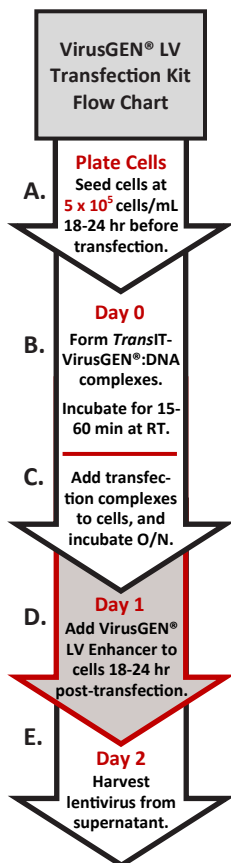
### SPECIFICATIONS

Storage	Store <i>TransIT-VirusGEN</i> ® Reagent at -10 to -30°C, tightly capped. <b>Before each use</b> , warm to room temperature and vortex gently. Store <i>VirusGEN</i> ® LV Complex Formation Solution and <i>VirusGEN</i> ® LV Enhancer at 2 to 10°C.
Product Guarantee	6 months from the date of purchase, when properly stored and handled.

### ► PROTOCOL FOR LENTIVIRUS GENERATION IN ADHERENT HEK 293 CELL CULTURES



Full protocol and additional documentation available at [mirusbio.com/virusgen](http://mirusbio.com/virusgen)



**Fill in volumes below based on culture vessel used for transfection (Table 1).**

#### A. Maintain cells

1. Approximately 18-24 hours prior to transfection, plate cells at a starting density of  $4 - 6 \times 10^5$  cells/ml. Cultures should be 80-95% confluent at the time of transfection.
2. Incubate cells overnight at appropriate temperature and CO<sub>2</sub> levels.

#### B. Prepare *TransIT-VirusGEN*® Reagent:DNA complexes

1. Warm *TransIT-VirusGEN*® Reagent to room temperature and vortex.
2. Place \_\_\_ ml of *VirusGEN*® LV Complex Formation Solution in a sterile tube.
3. Add \_\_\_ µl packaging DNA premix and \_\_\_ µl transfer plasmid DNA encoding the gene of interest. Mix gently by pipetting.
4. Add \_\_\_ µl of *TransIT-VirusGEN*® Reagent. Vortex gently to mix.
5. Incubate at room temperature for 15-60 minutes to allow transfection complexes to form.

#### C. Distribute complexes to cells in complete growth medium

1. Add *TransIT-VirusGEN*® Reagent:DNA complexes (from Step B) to cells.
2. Gently rock culture vessel for even distribution of complexes.
3. Incubate transfected cultures overnight.

#### D. Add *VirusGEN*® LV Enhancer to transfected cells

1. Between 18-24 hours post-transfection, add \_\_\_ mL of *VirusGEN*® LV Enhancer to cultures.
2. Incubate cultures for an additional 24-30 hours prior to virus harvest (i.e. harvest lentivirus a total of 48 hours post-transfection).

#### E. Harvest virus

1. Following the 48-hour incubation, harvest cell supernatant containing recombinant lentivirus particles.
2. Filter through a 0.45 µm PVDF filter to remove any cell debris.
3. Flash-freeze aliquots of lentivirus in cryotubes and store at -80°C.

**Table 1.** Recommended starting conditions

Culture Vessel	6-well plate	10-cm dish	T75 flask
Surface area	9.6 cm <sup>2</sup>	59 cm <sup>2</sup>	75 cm <sup>2</sup>
Complete growth medium	2.0 ml	10 ml	15 ml
LV Complex Formation Solution	200 µl	1.0 ml	1.5 ml
Packaging DNA Premix (1 µg/µl)	1 µl	5 µl	7.5 µl
Transfer Plasmid DNA (1 µg/µl)	1 µl	5 µl	7.5 µl
<i>TransIT-VirusGEN</i> ® Reagent	6 µl	30 µl	45 µl
<b>NOTE: Add <i>VirusGEN</i>® LV Enhancer 18-24 hours post-transfection.</b>			
<i>VirusGEN</i> ® LV Enhancer	200 µl	1.0 ml	1.5 ml

If using individual packaging plasmids, we recommend a starting ratio of 4 µg *gag-pol* vector, 1 µg *rev* vector and 1 µg VSV-G vector. Premix the packaging plasmids with an equal amount of the transfer vector (e.g. 6 µg) to maintain a 1:1 (wt:wt) ratio of packaging to transfer plasmids.

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### SPECIFICATIONS

Storage	Store <i>TransIT</i> -VirusGEN® Reagent at -10 to -30°C, tightly capped. <b>Before each use</b> , warm to room temperature and vortex gently. Store VirusGEN® LV Complex Formation Solution and VirusGEN® LV Enhancer at 2 to 10°C.
Product Guarantee	6 months from the date of purchase, when properly stored and handled.

### ► PROTOCOL FOR LENTIVIRUS GENERATION IN SUSPENSION HEK 293 CELL CULTURES



Full protocol and additional documentation available at [mirusbio.com/virusgen](http://mirusbio.com/virusgen)

#### Fill in volumes below based on total culture volume (Table 2).

##### A. Maintain cells

1. Passage suspension HEK 293 cells 18-24 hours prior to transfection to obtain a density of  $4 - 6 \times 10^6$  cells/ml the next day. Do NOT proceed with transfection if cells are not doubling every 24 hours or are < 95% viable.
2. Incubate cells overnight (e.g. 37°C, 5-8% CO<sub>2</sub>, shaking).

##### B. Prepare *TransIT*-VirusGEN® Reagent:DNA complexes

1. Immediately prior to transfection, seed cells at a density of  $4 \times 10^6$  cells/ml.
2. Warm *TransIT*-VirusGEN® Reagent to room temperature and vortex.
3. Place \_\_\_ ml of VirusGEN® LV Complex Formation Solution in a sterile tube.
4. Add \_\_\_ µl packaging DNA premix and \_\_\_ µl transfer plasmid DNA encoding the gene of interest. Mix completely.
5. Add \_\_\_ µl of *TransIT*-VirusGEN® Reagent. Vortex gently to mix.
6. Incubate for 15-60 minutes to allow transfection complexes to form.

##### C. Distribute complexes to cells in complete growth medium

1. Add *TransIT*-VirusGEN® Reagent:DNA complexes (from Step B) to cells.
2. Incubate transfected cultures overnight in an orbital shaker.

##### D. Add VirusGEN® LV Enhancer to transfected cells

1. Between 18-24 hours post-transfection, add \_\_\_ mL of VirusGEN® LV Enhancer to cultures.
2. Incubate cultures for an additional 24-30 hours prior to virus harvest (i.e. harvest lentivirus a total of 48 hours post-transfection).

##### E. Harvest virus

1. Following the 48-hour incubation, centrifuge cells in a sterile tube at 300 × g for 5 minutes. Do NOT dispose of the supernatant.
2. Transfer the virus-containing supernatant into a new sterile tube.
3. Filter through a 0.45 µm PVDF filter to remove any cell debris.
4. Flash-freeze aliquots of lentivirus in cryotubes and store at -80°C.

Table 2. Volume scaling worksheet for VirusGEN® LV Transfection Kit

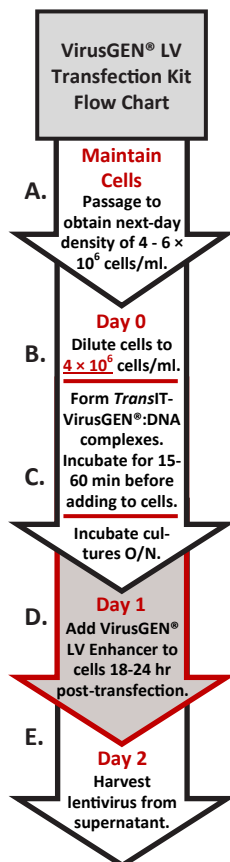
Starting conditions per milliliter of complete growth medium			
	Per 1 ml	Total culture volume	Reagent quantities
LV Complex Formation Solution	0.1 ml	× _____ ml	= _____ ml
Packaging DNA Premix (1 µg/µl)	0.5 µl	× _____ ml	= _____ µl
Transfer Plasmid DNA (1 µg/µl)	0.5 µl	× _____ ml	= _____ µl
<i>TransIT</i> -VirusGEN® Reagent	3 µl	× _____ ml	= _____ µl
<b>NOTE: Add VirusGEN® LV Enhancer 18-24 hours post-transfection.</b>			
VirusGEN® LV Enhancer	0.1 ml	× _____ ml	= _____ ml

If using individual packaging plasmids, we recommend a starting ratio of 4 µg *gag-pol* : 1 µg *rev* : 1 µg VSV-G vectors. Premix the packaging plasmids with an equal amount of the transfer vector (e.g. 6 µg) to maintain a 1:1 (wt:wt) ratio of packaging to transfer plasmids.

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