

Hygromycin B Solution

Quick Reference Protocol

Instructions for MIR 5930

MSDS and Certificate of Analysis available at mirusbio.com/5930



SPECIFICATIONS

Storage	Store Hygromycin B solution at 4°C. Protect from moisture.
Product Guarantee	Six months from date of purchase, when properly stored and handled.
Concentration	50 mg/ml Hygromycin B Free Base, sterile filtered in PBS
Handling	TOXIC. Handle with care. See MSDS for more information.

▶ ANTIBIOTIC KILL CURVE PROTOCOL



MSDS and Certificate of Analysis available at mirusbio.com/5930

Hygromycin B antibiotic ensures effective positive selection of cells expressing the hygromycin resistance (*hph*) gene. In mammalian cells, the recommended working concentration range for hygromycin B is 100 – 500 µg/ml. Different cell types and cell culture conditions may require different concentrations of selection antibiotic. Perform a kill curve to determine the optimal working concentration for your experiment. The following is a general guideline for performing an antibiotic kill curve.

NOTE: Performing a kill curve is recommended with each new cell type or selection antibiotic lot, or if changes are made to the cell culture conditions.

- A. Plate cells in 0.5 ml complete growth medium per well in a 24-well tissue culture plate.
 - For adherent cells:** Plate cells at a density of $0.8\text{--}3.0 \times 10^5$ cells/ml.
 - For suspension cells:** Plate cells at a density of $2.5\text{--}5.0 \times 10^5$ cells/ml.
- B. Culture overnight. Most cell types should be $\geq 80\%$ confluent prior to adding the selection antibiotic.
- C. Add increasing amounts of hygromycin B to duplicate wells of cells plated in complete media. Include a no-antibiotic control. For example, add 0, 25, 50, 100, 150, 200, 250, 300, 400, and 500 µg/ml hygromycin B to duplicate wells of cells plated in complete growth media. Certain cell types and cell culture conditions may require concentrations outside of this range.
- D. Replace media containing selection antibiotic every 2-3 days for up to a week. Examine the culture every day for signs of visual toxicity. Determine the following antibiotic doses:
 - **Low dose** - the antibiotic concentration at which minimal visual toxicity is apparent after 7 days of antibiotic selection
 - **Optimal dose** - the lowest antibiotic concentration at which all cells are dead after 7 days of antibiotic selection
 - **High dose** - the antibiotic concentration at which visual toxicity is evident within the first 2-3 days of antibiotic selection
- E. Proceed with stable cell line generation using the concentrations determined in step D. Cells transfected with a plasmid harboring the hygromycin resistance (*hph*) gene should be grown in complete growth medium for 48–72 hours post-transfection before selection antibiotic is applied. For more information on stable cell line generation, visit www.mirusbio.com/stable.

▶ NOTES

©1996-2018 All rights reserved. Mirus Bio LLC. All trademarks are the property of their respective owners.
For terms and conditions, visit www.mirusbio.com

Rev.A 1216